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WITNESS my hand this
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PROVISIONAL SPECIFICATION

Applicant(s):

COMMONWEALTH SCIENTIFIC AND INDUSTRIAL RESEARCH ORGANISATION

Invention Title:

METHOD FOR MICROBIAL DISCOVERY

The invention is described in the following statement:

METHOD FOR MICROBIAL DISCOVERY

FIELD OF THE INVENTION

The present invention relates to a method for microbial discovery. In particular, the present invention relates to a method for selectively enriching and thereby discovering a microorganism which can metabolise a test substrate. The present invention further relates to an apparatus for conducting the method and microorganisms discovered by the method.

BACKGROUND OF THE INVENTION

Techniques such as passaging in batch culture are still used today for the discovery of microorganisms which can metabolise a test substrate. These techniques are often labour intensive, slow and the expected outcome is not known until the enriched microbial population is plated onto selective media. Traditional methods for monitoring the activity or growth of a microbial population include measurement of biomass concentration and/or measurement of substrate consumption. These analytical techniques do not provide an assessment of the status of a microbial population in real-time to enable the status of the microbial culture to be determined, and intervention to occur if necessary.

The chemostat provides continuous culture and has been used for enrichment to facilitate the discovery of microorganisms with useful properties and the study of evolutionary pathways. The effectiveness of conventional continuous culture is limited because the status of the discovery process cannot be evaluated rapidly. In a limited number of cases carbon dioxide production and oxygen consumption have been used to monitor a continuous culture. However these techniques have been subject to a range of limitations imposed by the small number of applications to which the techniques have been considered applicable, and/or limitations based on deficiencies in

the apparatus. For one example, off-line analysis of biomass concentration or residual substrate concentration is commonly required to evaluate the status of an enrichment process. Off-line analysis is time consuming in terms of the slow analytical techniques involved, and indeed the delays in developing an appropriate analytical procedure for determining analyte concentration. Furthermore, a significant level of infrastructure and staff trained in the use of the analytical equipment are also required.

Therefore, the applicants have identified a need for faster methods for microbial discovery, and apparatus therefor, particularly for use in a new application identified by the applicants.

SUMMARY OF THE INVENTION

Accordingly, the present invention provides a method for assessing the selective enrichment of a microorganism able to metabolise a test substrate, the method comprising the steps of

a) providing a population of microorganisms in a vessel,

b) b) feeding fluid into the vessel at a controlled flow rate commencing with an initial flow rate, the fluid comprising a nutrient medium and, for at least part of the feed period, the test substrate,

c) providing a probe for producing a signal indicative of the level of a metabolism indicator over the time-frame of the enrichment, and

d) providing an output based on the signal to enable assessment of selective enrichment of a microorganism that metabolises the test substrate.

The present inventors have found that the on-line determination of a change in the level of a metabolism indicator, such as O_2, as an indicator of cellular activity enables indirect measurement of biomass or substrate utilisation and have identified that this can be used to

evaluate the status of a population of microorganisms in real-time. The inventors have further tailored this technique for enriching microorganisms that are capable of metabolising a test substrate, such as a hydrocarbon compound for which a microorganism is desired to be found to convert the compound into a different hydrocarbon(s) and/or break the compound down with water as a byproduct.

The technique developed by the inventors has further advantages in terms of its flexibility in discovering microorganisms capable of metabolising a test substrate in conditions selected by the operator, and potentially modified by the operator over time. All of this is evaluated in real-time without the need to separately measure substrate levels or determine biomass concentration.

In a preferred embodiment, the method further comprises presetting conditions to be met by the signal output to result in a change in the fluid flow rate, and changing the flow rate at which fluid is fed into the vessel when the conditions are met, wherein the preset conditions are a predetermined period of time, and a preset value range within which the signal must remain for the predetermined period of time.

The flow rate of the fluid fed into the vessel is suitably increased from the initial flow rate on meeting the preset conditions to reduce the hydraulic retention time, and thereby increase selectivity for a microorganism that metabolises the test substrate. Increasing the flow rate of the fluid fed into the vessel will facilitate the selective enrichment of microorganisms which metabolise the test substrate more quickly and therefore reproduce more quickly. In effect, the preset conditions should be set to define the maintenance of steady state in the culture over the predetermined time period. The predetermined time period may be in a time unit of measurement (eg a number of minutes or hours), or may be set by reference to a predetermined multiple (including

fractions) of the hydraulic retention time of the vessel. Consequently it will be understood that the reference to a predetermined time period need not be an exact, repeated number of hours, especially if the fluid flow rate is changed over time.

The flow rate of the fluid fed into the vessel may be increased by increasing the flow rate of the test substrate. Further, the fluid flow rate may be increased by increasing the flow rate of the nutrient medium in addition to the test substrate. If the level of test substrate in the vessel is sufficiently high it is possible for the flow rate to be increased by increasing the flow rate of the nutrient medium alone, although this is not preferred. Where the flow rate of both the test substrate and the nutrient medium is increased, it is convenient for the flow rates to be increased proportionally such that the concentration of the test substrate in the fluid fed into the vessel remains substantially constant.

The metabolism indicator used in the method of the invention may be the uptake or release of a molecule involved in metabolism of the test substrate. According to one embodiment, the metabolism indicator is selected from the group consisting of oxygen, carbon dioxide, sulfate, nitrate and iron. Preferably the metabolism indicator is selected from oxygen, sulphate, nitrate and iron, and most preferably it is oxygen.

The signal of the level of the metabolism indicator is preferably provided as a visual output, such as a plot of points which represent the level of the metabolism indicator against time. The signal output of the probe will be an electrical signal, and therefore the plot may be of the electrical output (eg current) against time. Otherwise, in the example of the metabolism indicator being oxygen uptake, the electrical signal may be converted into oxygen concentration or oxygen uptake rate, and this may be plotted against time. The output could

also be a numerical digital or liquid crystal display. The visual output may conveniently be updated in periods of less than 20 minutes. Ideally, the visual output is updated in periods of 10 mins or less.

The population of microorganisms used in the method of the invention may be a heterogeneous population, such as activated sludge, or may be a homogeneous population.

The method of the invention may further comprise the step of subjecting the population of microorganisms to a mutagen, such as a chemical mutagen or ultra-violet light. The method of the invention may further comprise the step of isolating the enriched microorganism.

The present invention further provides a microorganism when enriched or isolated by the method described above.

According to the present invention, there is also provided an apparatus for the selective enrichment from a population of microorganisms of a microorganism able to metabolise a test substrate, the apparatus comprising:

(a) a vessel for receiving the population of microorganisms, a nutrient medium and the test substrate;

(b) a probe for producing a signal indicative of the level of a metabolism indicator in the vessel;

(c) a supply mechanism for supply of the nutrient medium and the test substrate to the vessel; and

(d) a controller for changing operation of the supply mechanism for at least one of the nutrient medium and the test substrate in response to the probe signal being within a preset range for a predetermined period of time, to thereby enrich for microorganisms capable of metabolising the test substrate.

It will be understood from the following that the probe signal may be converted into another unit relating to the level of a metabolism indicator being measured by the probe. It will be understood therefore that the preset range for the probe signal may be set as a direct signal value, or indirectly by reference to the level of

the metabolism indicator, or any other related unit of measurement. Thus, according to one embodiment, the preset range for the probe signal is set by reference to the level of the metabolism indicator which is represented by the probe signal.

In most situations, the controller will be set to increase the flow of nutrient medium and/or test substrate into the vessel in response to the probe signal meeting the preset conditions. This particularly enables the apparatus to select for microorganisms that metabolise the test substrate and reproduce quickly, as microorganisms not able to reproduce quickly enough will be washed out of the apparatus. Thus, according to one embodiment, the supply mechanism operates to supply the nutrient medium and the test substrate to the vessel at an initial flow rate, and the controller is set to increase the flow rate from the initial flow rate in response to the probe signal meeting the preset conditions. However, it is appreciated by the applicants that decreases could be set, especially in a later stage of an operation being conducted on the apparatus.

Generally, the intention of pre-setting the range (upper and lower signal ranges) of the probe signal is to identify when the culture has reached a steady-state. Once a steady state has been identified, it is possible to change the flow of fluid (nutrient medium and/or test substrate) into the vessel.

The supply mechanism of the apparatus may operate to supply a mixture of test substrate and nutrient medium into the vessel, in a known ratio, or the supply mechanism may operate to supply the test substrate and nutrient medium separately. Many advantages flow from independent supply mechanisms. Firstly, being able to supply the two fluids separately offer more control to the user in terms of modifying the conditions under which the microorganisms are required to metabolise and reproduce. Secondly, this offers advantages in terms of switching from one test

substrate to the next without changing the nutrient medium fed into or supplied by the nutrient medium supply mechanism.

It is possible for the preset range of the probe signal being surveyed by the controller to be preset into the apparatus. However, according to a particularly useful embodiment, the controller comprises a user interface which enables the user to set the upper and lower limits of the range. In the case where the probe is producing a signal representative of the level of oxygen in the vessel, the controller may allow the user to select the maximum and minimum levels in any appropriate unit of measurement, such as ml/l of oxygen to liquid in the vessel, biological oxygen demand (BOD), oxygen uptake rate (OUR) or similar. Of course, where the metabolism indicator being detected by the probe is another indicator such as carbon dioxide, nitrate, iron or so forth, the controller may allow the user to select the maximum and minimum levels in the unit of measurement relevant to those signals.

As indicated above, the probe signal must be within a preset range for a predetermined period of time before the controller will respond. The predetermined period of time may also be preset for the apparatus, however it is again preferred that the user interface enables the user to set this time period.

Preferably the apparatus further comprises a display for providing a visual output of the probe signal over time and/or the level of other conditions existing in the vessel. The conditions which may be displayed include the pH, temperature, aeration level and so forth.

Preferably, the user interface enables the user to set the pH level and temperature of the vessel, the supply means controls the supply of chemicals necessary to alter the pH, the apparatus includes temperature control means, and the controller controls the supply means and the temperature control means in response to changes made to

the pH and temperature levels set through the user interface. As will be understood, the provision of this function operated by the user enables the user to modify the conditions to select a microorganism able to metabolise the test substrate in specific conditions (eg high or low pH; high or low temperature etc). Ideally, the pH and/or temperature controller operate to increase or decrease the pH and/or temperature incrementally in increments over a time period that can be set by the user.

The user interface may further enable the user to set other conditions that impact on the metabolism, such as the oxygen level or aeration rate.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a schematic illustration of the apparatus of one embodiment of the invention.

Figure 2 is a schematic illustration of the apparatus of Figure 1 with further apparatus components.

Figure 3 shows the correlation between OUR and microbial activity as determined by conventional analytical techniques, as well as the correlation between different conventional analyses, using acetic acid as the test substrate.

Figure 4 shows the correlation between OUR and microbial activity as determined by conventional analytical techniques, as well as the correlation between different conventional analyses, using sodium acetate as the test substrate.

Figure 5 shows the correlation between OUR and microbial activity as determined by conventional analytical techniques, as well as the correlation between different conventional analyses, using benzyl alcohol as the test substrate.

Figure 6 demonstrates the correlation between a population change and BOD - the BOD and residual substrate concentration.

Figure 7 demonstrates the correlation between a population change and BOD - the changes to the population as measured using viable cell counts and optical density.

Figure 8 shows the increase in BOD after the addition of 1-methyl-2-pyrrolidinone to a culture.

Figure 9 shows BOD during growth of microorganisms from activated sludge on 1-methyl-2-pyrrolidinone.

Figure 10 shows BOD output during growth of microorganisms from activated sludge using dodecane as the test substrate.

Figure 11 shows the output of the control panel screen for monitoring enrichment in real-time, with feedback loop between feed flow rate and BOD added.

Figure 12 shows the effect of flow rate on the BOD of a 1,3-propanediol-degrading microbial population.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

The present invention provides a method for assessing the selective enrichment of a microorganism able to metabolise a test substrate. It will be understood that a "microorganism" means any microorganism, for example, bacteria, fungi, yeast, protozoans, algae or viruses. Any of these microorganisms can be selectively enriched by designing the enrichment conditions to favour the growth of a microorganism with a particular characteristic.

It must be noted that as used herein, the singular forms "a", "an", and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, reference to a microorganism includes a plurality of microorganisms.

As used herein, the term "enrichment" means an increase in the number of microorganisms in a population which are able to metabolise the test substrate. "Metabolise" means to use the test substrate in a chemical reaction within the microorganism by either catabolism or anabolism. Therefore a test substrate may be used in a chemical reaction that combines the test substrate into a

more complex molecule, or may be used in a chemical reaction which breaks down the test substrate into a simple molecule.

The "test substrate" is any substrate for which it is desired to screen for a microorganism able to metabolise the test substrate and does not include substrates which are commonly metabolised, such as glucose. The purpose of the method of the invention is to arrive at a microorganism population that is able to metabolise the test substrate. Therefore the technique and the controls required are very different to techniques where the substrate is known to be a substrate for certain microorganisms, or is a common substrate for a large range of microorganisms. Typically, the method of the invention will be used to selectively enrich microorganisms which can metabolise an organic carbon-containing molecule. The term "organic carbon-containing molecule" refers to aliphatic and aromatic hydrocarbons and derivatives thereof, including carbohydrates other than commonly metabolised substrates such as glucose. Alternatively, the method of the invention may be used for the enrichment of microorganisms capable of anaerobic respiration metabolising for example, sulphur-containing metabolism indicators or nitrogen-containing metabolism indicators.

The method comprises the step of providing a population of microorganisms in a vessel. It will be clearly understood that the population of microorganisms may be a homogeneous population of microorganisms or may be a heterogeneous population of microorganisms. A homogeneous population may be useful to selectively enrich for a microorganism by evolution. Where the population of microorganisms is a heterogeneous population this may be, for example, a microbial library or a heterogeneous population such as activated sludge. Activated sludge is the product that results when primary effluent of raw sewage is mixed with bacteria-laden sludge and then agitated and aerated to provide biological treatment in

order to speed the breakdown of organic matter in the raw sewage undergoing secondary waste treatment. The present inventors have successfully used activated sludge as the starting microbial population in the method of the invention to enrich for microorganisms able to metabolise a large number of diverse substrates under a diverse range of conditions.

The fluid comprises a nutrient medium and the test substrate. A "nutrient medium" is a growth medium which comprises all of the nutrients required for growth of a microorganism but essentially no amount of the test substrate or substrates similar to (eg in the same class as) the test substrate. The concept of "Similar substrates" to the test substrate is described below. The nutrient medium will depend upon the microbial population being enriched and the substrate being tested. However it is generally a nitrogen (ammonium), phosphorus, sulphur, salt (eg Na, Mg, Ca) and trace metal-containing solution. For example, when the method of the invention is used to test the ability of *Pseudomonas putida* F1 to metabolise acetic acid, the nutrient medium may be that set out in the Examples below. The nutrient medium may contain a trace amount of the similar substrate provided that the amount does not interfere with the detection of the enrichment process. The amount of the similar substrate must be such that it does not interfere with detection of the enrichment process. Ideally, the nutrient medium contains no similar substrates. For example, where the test substrate is an organic carbon-containing test substrate the nutrient medium contains substantially no organic carbon-containing material. There is also the possibility that the test substrate could be used as the sole source of another nutrient other than carbon, for example nitrogen or sulphur. In this case the nitrogen or sulphur would need to be eliminated from the nutrient medium or kept at a concentration that does not interfere with enrichment process.

"Similar substrate" means a substrate which the microorganism can metabolise as an alternative to the test substrate. For example, where the method is used to selectively enrich a microorganism able to catabolise a particular organic carbon-containing substrate, a similar substrate is an alternative carbon-containing substrate which the microorganism is able to catabolise. Where the test substrate is a small hydrocarbon molecule, "similar substrates" to be avoided in the nutrient medium are other small hydrocarbon (including carbohydrate) molecules, such as glucose.

The test substrate may be fed into the vessel as part of the nutrient medium or separately to the nutrient medium. For better control, these fluids can be fed into the vessel independently. The initial flow rate at which the nutrient medium and test substrate are fed into the vessel, or hydraulic retention time, is chosen by reference to factors such as the starting population of microorganisms, the nutrient medium, the temperature of the vessel and the fluid, the pH of the fluid, and the stage of enrichment, and the vessel volume. Hydraulic retention time is a measure of the length of time that liquid remains in the vessel. It equals V/Q (V = vessel volume, Q = flow rate). Typically the initial hydraulic retention time will be relatively long in order to establish a steady state within the vessel.

The selective enrichment of a microorganism is made possible through the on-line monitoring of the level of metabolism indicator with the probe and the real-time output based on the probe signal.

As used herein, "on-line" means that the probe is in direct contact with the contents of the vessel, be that the fluid in the vessel or gas in the headspace of the vessel, the probe taking readings in the vessel itself or in a conduit through which contents of the vessel may flow. The purpose of this arrangement is to enable signal readings to be taken without removal of fluid from the

apparatus, including the vessel and any associated conduits. Monitoring the level of a metabolism indicator on-line alleviates the need for off-line analyses in order to monitor enrichment and therefore facilitates the real-time determination of enrichment.

As used herein, "real-time" means that the output of the level of the metabolism indicator is provided fast enough to enable the status of the microbial culture in response to a change in conditions to be determined, and intervention to occur if necessary. An example of intervention provided by real-time monitoring is that which prevents the loss of a microbial population in response to a change in the conditions of the population that does not enable metabolism of the test substrate by a microorganism in the population. The frequency required to provide the output of the level of the metabolism indicator will depend upon the status of the enrichment process and the growth rate of the microorganism being enriched. The output of the level of the metabolism indicator should be updated in periods of 20 minutes or less.

The metabolism indicator may be any indicator of metabolism, for example a molecule consumed during metabolism such as oxygen, or a molecule produced by metabolism, provided only that the level of the metabolism indicator is able to be monitored on-line and used to provide an output of the level of the metabolism indicator. The metabolism indicators identified as being capable of being monitored on-line with a probe are oxygen, carbon dioxide, sulphate, nitrate and iron. These molecules act as terminal electron acceptors in the metabolism and probes are available for detecting levels of these molecules in solution.

According to one embodiment, the oxygen uptake rate (OUR) of the microbial culture may be used as the metabolism indicator, particularly for the identification of aerobes. This can be determined by adding oxygen to the

culture followed by the determination of a change in the oxygen level after a specific time period. The OUR gives a real-time measure of both substrate utilisation and growth of the population. By using this value to calculate the biological oxygen demand (BOD) of the test substrate in the fluid fed into the vessel, the level of substrate utilised can be determined. This is described further below in the examples.

Similar calculations can be used for any other metabolism indicator and probe combination. For example, in the situation where the microbe is an anaerobe and does not use oxygen to respire during metabolism of the target molecule, but instead uses sulphate as the terminal electron acceptor, a sulphate probe can be used to monitor levels of sulphate.

The method of the invention may further comprise subjecting the microorganism population to a mutagen. As used herein, a mutagen is an agent which induces a change in the phenotype of a microorganism. A person skilled in the art will be readily able to determine a suitable mutagen, for example a chemical mutagen or an ultra-violet light with a wave length of 10 nm to 400 nm may be used.

The method of the invention may further comprise the step of discovering the enriched microorganism. That is, isolating the enriched microorganism. This step may be readily performed by the person skilled in the art using standard microbiological techniques. For example, where the enriched microorganism is a bacteria, a sample of the enriched culture may be plated onto solid nutrient medium which contains the test substrate, and the plate incubated under the conditions which enable enriched bacteria to metabolise the test substrate. Individual colonies formed by the enriched bacteria can then be isolated, and subjected to further characterisation steps if required.

Where not otherwise described herein, the techniques employed in putting the invention into practice are conventional microbiological and chemical techniques known

within the art. Such techniques are well known to the skilled worker, and are fully explained in the literature. See, eg., Bergey's Manual of Systematic Bacteriology; Bergey's Manual of Determinative Bacteriology; The Prokaryotes, Starr, Stolp, Truper, Balows, Schlegel, editors; Handbook of Microbiological Media, Atlas; Biology of Microorganisms, Brock, Madigan, Martinko and Parker; Methods for General and Molecular Bacteriology, Gerhardt, Murray, Wood, Krieg, editors.

The invention will now be described by way of the following non-limiting examples and drawings. Although any materials and methods similar or equivalent to those described herein can be used to practice or test the invention, the preferred materials and methods are now described.

EXAMPLES

Figures 1 and 2 illustrate an example of the apparatus of one embodiment of the invention. The apparatus comprises a vessel or bioreactor 1 with an oxygen (air) injection means 2 and a dissolved oxygen measuring probe 3. The vessel is also associated with a temperature control means, including a temperature probe 4. The vessel also includes a stirrer 5 for stirring the contents of the vessel.

Fluid is fed into the vessel through inlet 6. The embodiment illustrated contains one inlet for feeding a combination of nutrient medium and test substrate, however separate inlets for each may be provided. A supply mechanism (not illustrated) controls flow of fluid into the vessel via inlet 6. The supply mechanism is connected to a nutrient medium supply well and a test substrate supply well (also not shown) to enable the control of the ratio of the two fluids, and the flow rate into the vessel 1. Overflow fluid is removed from the vessel via fluid outlet 7.

The apparatus further comprises an inlet 8 for the supply of acid and alkali for the control of pH in the vessel. The pH of the fluid in the vessel is measured by a pH probe 9.

Further components of the apparatus illustrated include electronics plugs 13, a sample line/drain 14 and a screw collar 15. Numeral 16 refers to the vessel base and 17 to the top of the vessel/stirrer.

The apparatus according to the embodiment illustrated may be provided as a unit 10 containing the elements described above, together with a control unit 11. The control unit 11 of this embodiment is under the control of a computer 12, which includes a monitor and a keyboard. The computer is programmed to provide a graphical user interface with the control program which allows the user to control the parameters described in the Examples that follow. The computer interacts with the control unit so that they together operate to control the supply mechanism to control the supply of fluids into the vessel in response to the probe signal.

The apparatus of the embodiment illustrated provides a series of visual outputs, including the screen output illustrated in Figure 11. This output shows the settings entered by the user for defining the pH, temperature, aeration level, upper and lower limits of the probe signal range (measured in this case in terms of the level of oxygen, measured in mg l^{-1}), the initial flow rate of inlet fluid, the flow increment (positive value represents increase), and the predetermined time period (which can be set as a number of vessel volumes).

From this screen output, the screen can be switched to an output of one of a number of graphs including those illustrated (with entries) in Figures 6 to 10 and 12.

The mechanical and program components of the apparatus will be well understood to those skilled in the relevant arts, in the light of the functional description provided herein.

In the following examples, the nutrient medium used was a defined medium (DM) prepared as outlined in Appendix 1.

EXAMPLE 1: CORRELATION BETWEEN OXYGEN UPTAKE RATE (OUR) AND MICROBIAL ACTIVITY

To determine whether the oxygen uptake rate (OUR) is a true reflection of the activity of a microbial population, OUR was compared with analytical techniques that are typically used to evaluate microbial activity. A 100 ml shake flask culture of *Pseudomonas putida* F1 (ATCC 70007) that had been grown for 48 hours at 28°C shaking at 190 rpm, then centrifuged and resuspended in 10 ml of defined medium (DM) with no carbon source added, was used to inoculate DM that contained 1.5 or 2.0 g l^{-1} acetic acid or 1.0 g l^{-1} benzyl alcohol. After inoculation the culture was sampled periodically for determination of microbial activity by conventional analytical techniques such as viable cell number, optical density (600 nm) and residual substrate concentration. OUR was measured every 10 minutes. These conventional analyses were compared with OUR measured using the method of the invention. The experiment was repeated three times, twice with acetic acid as the test substrate and once with benzyl alcohol. The correlated results of the three experiments are shown in Figures 3, 4 and 5.

From the data shown in Figures 3, 4 and 5 there is a clear linear correlation between OUR and both substrate consumption and biomass concentration regardless of whether the substrate is acetic acid or benzyl alcohol. The correlation between biomass and substrate utilisation shows a clear exponential correlation. This is probably because the yield value ($Y_{x/s}$; grams of biomass per gram of substrate) is not a true constant and is actually dependent on growth rate which is changing constantly during growth in batch culture (Mandelstam et *al.*, Biochemistry of Bacterial Growth. 3rd Edition,

Blackwell Scientific Publications, Oxford, UK, 1982). The method of the invention can therefore be used as a superior alternative to monitor the status of enrichment in real-time. This provides the operator with the opportunity to rapidly refine the culture conditions or determine the effect on a culture of changing the many parameters which can affect the enrichment of a microbial population.

EXAMPLE 2: DEMONSTRATION OF REAL-TIME MONITORING OF A POPULATION CHANGE

To test and demonstrate the operation of the method of the invention, a control experiment was performed which compared the output of the method with off-line measurements that are traditionally used to monitor microbial activity. Techniques that are typically used include measurement of the residual substrate concentration and/or measurement of biomass concentration (viable count and optical density). These methods were compared with the output of the present method to demonstrate the utility of the method.

For these control experiments a steady state culture of an *Escherichia coli* BL21DE3 which was supplied by Novagen (Novagen Inc., Madison, WI, USA) and was expected to grow on glucose only was used. The culture was established using 5 ml of an E. coli culture taken from a 100 ml shake flask culture which had had been grown for 17 hours shaking at 200 rpm and 30°C in defined medium with 1.0 g l^{-1} glucose as the carbon source. Although the feed contained another substrate (benzyl alcohol), no growth on this substrate was expected because this microbial population was known to be unable to use this carbon source for growth. When steady state had been established, 10 mL of a 100mL shake flask culture of *Pseudomonas putida* F1 was added. The *P. putida* F1 culture had been grown at 30°C for 17 hours, with shaking at 200 rpm, in defined medium with 1.0 g l^{-1} glucose as the carbon

source. The *P. putida* F1 was supplied by the American Type Culture Collection (ATCC) and was expected to grow on benzyl alcohol and/or glucose. The OUR was expected to change as a result of the increased microbial activity after the addition of *P. putida*.

2.1 Growth of *E. coli* and *P. putida* on and in the presence of glucose and benzyl alcohol in defined medium in batch culture

The success of this control experiment was dependent upon the ability of *E. coli* to grow on glucose in DM and grow in DM in the presence of benzyl alcohol (*i.e.* benzyl alcohol is not toxic to *E. coli*). Also of key importance was the inability of *E. coli* to grow on benzyl alcohol. Similarly, it was important to demonstrate growth of *P. putida* on benzyl alcohol. Although *P. putida* is well known for its ability to grow on a wide range of aromatic substrates (Wackett, & Hershberger, 2001), growth on benzyl alcohol has not been reported. The ability of each of the two strains to grow under the conditions used in the method is shown in Table 1. The optical density at inoculation was calculated (based upon the optical density on the inocula) as 0.021 (*E. coli*) and 0.026 (*P. putida*). The cultures were incubated shaking at 200 rpm and 30°C. The optical density was measured at 600 nm after incubation for 23.5 and 75 hours.

Table 1: Growth of *E. coli* and *P. putida* on glucose and benzyl alcohol in batch culture.

Carbon source		Organism (Optical density at 600 nm after 23.5 and 75 hours incubation)			
Glucose (0.1 g l^{-1})	Benzyl alcohol (1.0 g l^{-1})	*E. coli* 23.5 hours	*E. coli* 75 hours	*P. putida* 23.5 hours	*P. putida* 75 hours
Not added	Not added	0.026	0.027	0.050	0.046
+	Not added	0.116	0.102	0.170	0.138
+	+	0.102	0.097	0.042	0.353
Not added	+	0.027	0.024	0.021	0.417

From the data shown in Table 1 it is clear that *E. coli* can grow on glucose in DM and cannot grow on benzyl alcohol in DM, but will grow in the presence of 1.0 g l^{-1} benzyl alcohol. It is important that *E. coli* is able to tolerate benzyl alcohol as this will be present in the feed fluid for the entire experiment. It was established that *P. putida* was able to grow in DM containing both glucose and benzyl alcohol.

2.2 Measurement of a characterised population shift using the method of the invention

DM was inoculated with *E. coli* to give a starting optical density (measured at 600 nm) of 0.06 and then operated in batch mode for 19 hours during which time the BOD increased to approximately 200 mg l^{-1}. The BOD then declined rapidly indicating that the glucose in the medium was exhausted. When the fresh medium was pumped into the vessel the BOD increased again, peaking at just over 200 mg l^{-1} before stabilising at ~185 mg l^{-1}. Based on the calculated BOD for a feed fluid containing 0.5 g l^{-1} glucose the BOD was expected to be 178 mg l^{-1} (see calculation below).

Balance the stoichiometry of the following equation:

$$C_6H_{12}O_6 + O_2 \rightarrow CO_2 + H_2O$$

i.e.

$$C_6H_{12}O_6 + 6O_2 \rightarrow 6CO_2 + 6H_2O$$

Therefore:

Complete oxidation of 1 mol of $C_6H_{12}O_6$ requires 6 mol of O_2

Convert from moles to grams:

180.2 grams of $C_6H_{12}O_6$ requires 32 x 6 grams O_2

180.2 g of $C_6H_{12}O_6$ requires 192 g of O_2

Concentration of glucose in the feed = 0.5 g l^{-1}, therefore:

0.5 g of $C_6H_{12}O_6$ requires 0.53 g of O_2

Therefore the Chemical Oxygen Demand (COD):

COD = 530 mg l^{-1}

The BOD is assumed to be one third of the COD:

BOD = 178 mg l^{-1}

The correction factor for conversion of COD to BOD was determined experimentally using acetate as the carbon

source. The BOD of a known concentration of acetate was determined experimentally and compared to the calculated COD for the same concentration of acetate and the difference was found to be three-fold. It is assumed that the same conversion factor can be used for a range of readily biodegradable substrates.

The actual BOD was slightly higher than the calculated BOD for the substrate due to background respiration of the culture. Background respiration can be attributed to maintenance energy production and is therefore dependent upon the biomass concentration in the reactor. As the substrate concentration was relatively low the biomass concentration was also low and similarly the background respiration was low. Background respiration can be determined after the culture has reached steady state. The feed fluid flow is reduced to 0 ml h^{-1} and a rapid decrease in BOD is observed. Despite the absence of any readily degradable carbon the BOD is usually greater than zero. After a period of stabilisation the BOD will attain a steady value which is an indication of the background respiration.

Based on the BOD it was clear that steady state had been attained (it is generally assumed that steady state has been established after turnover of a least three vessel volumes which, in this example would occur after 37.5 hours). After 125.7 hours (equivalent to 10 vessel volumes) of continuous operation, *P. putida* was added to the culture. Initially, there was no change in the BOD so to ensure *P. putida* was not being washed out of the vessel the feed flow rate was reduced from 60 ml h^{-1} to 30 ml h^{-1}. The BOD increased slowly indicating that degradation of benzyl alcohol was beginning to occur. This observation was confirmed by measuring the residual benzyl alcohol in the culture supernatant, which had started to decrease. As the *P. putida* population developed the BOD increased, peaking initially at nearly 1400 mg l^{-1} before declining to 1050 mg l^{-1} after which a second peak in the BOD was

observed. The reason for the oscillation in the BOD is not clear although before reaching steady state microbial populations can demonstrate oscillations as the system equilibrates. After the second BOD peak, the BOD stabilised at 1040 mg l^{-1} which was the expected BOD for a feed containing 0.5 g l^{-1} glucose and 1.0 g l^{-1} benzyl alcohol (see calculation below).

Balance the stoichiometry of following equation:

$$C_7H_8O + O_2 \rightarrow CO_2 + H_2O$$

i.e.

$$2C_7H_8O + 17O_2 \rightarrow 14CO_2 + 8H_2O$$

Therefore:

Complete oxidation of 2 mol of C_7H_8O requires 17 mol of O_2

Convert from moles to grams:

108.1 x 2 grams of C_7H_8O requires 32 x 17 grams O_2

216.2 g of C_7H_8O requires 544 g of O_2

Concentration of benzyl alcohol in the feed = 1.0 g l^{-1}, therefore:

1.0 g of C_7H_8O requires 2.52 g of O_2

Therefore the Chemical Oxygen Demand (COD):

COD = 2516 mg l^{-1}

The BOD is assumed to be one third of the COD:

$$BOD = 839 \text{ mg } l^{-1}$$

As the feed contains both 0.5 g l^{-1} glucose and 1.0 g l^{-1} benzyl alcohol expected Evolver output when both substrate are being used by the microbial population in the reactor:

$$BOD = 839 + 178 = 1017 \text{ mg } l^{-1}$$

The feed comprised of DM containing 0.5 g l^{-1} glucose and 1.0 g l^{-1} benzyl alcohol was fed into a vessel at 30°C and pH 7. The feed flow rate was initially 60 ml h^{-1}. As shown in Figure 6, after inoculation of the reactor with *E. coli* (*Arrow A*), a population of microorganisms which could only use glucose as a carbon source for growth was established (*Arrow B*). *P. putida*, which can use benzyl alcohol as a carbon source for growth, was then added to the reactor (*Arrow C*) and the feed flow rate was reduced to 30 ml h^{-1} (*Arrow D*). A resultant increase in BOD and decrease in benzyl alcohol concentration were observed (*Arrow E*). The residual benzyl alcohol concentration was estimated using gas chromatography. At the same time that the BOD stabilised the measured residual benzyl alcohol concentration was zero. Interestingly, with a feed flow rate of 30 ml h^{-1} steady state was expected to be attained after 75 hours. However, based on the BOD, steady state that was not achieved until 94 hours after the feed flow rate was reduced from 60 ml h^{-1} to 30 ml h^{-1}. From this observation the microbial discovery process will be improved by waiting at least four vessel volumes before assuming a microbial population has reached steady state.

During the course of the experiment the biomass concentration was also monitored as was the number of benzyl alcohol-degrading microorganisms in the population (Figure 7). Viable cell numbers were estimated by plating samples of the culture (diluted in DM with no added carbon

source) on to solid DM containing either 1.0 g l^{-1} glucose or 1.0 g l^{-1} benzyl alcohol. The optical density of the culture was measured at 600 nm; samples were diluted in water if the optical density was greater than 0.4. After inoculation of the reactor with *E. coli* (Arrow A), a population of microorganisms which could only use glucose as a carbon source for growth was established. *P. putida*, which can use benzyl alcohol as a carbon source for growth, was then added to the reactor (Arrow B). A resultant increase in optical density, the total number of viable cells and the number of cells that could grow on benzyl alcohol, was observed (Arrow C). The observed increase in biomass concentration (Figure 7) correlated with the increase in BOD shown in Figure 6. The *E. coli* population growing on glucose and at steady state contained 2.52×10^9 cfu (colony forming units) ml^{-1} of culture, none of which could grow on benzyl alcohol. The inability of the *E. coli* population to grow on benzyl alcohol was confirmed by plating undiluted culture onto defined medium with benzyl alcohol as the only carbon source. Immediately after addition of *P. putida* to the culture the number of microorganisms growing on benzyl alcohol increased to 4.47×10^6 cfu ml^{-1}. In parallel with the increase in the BOD, the number of microorganisms in the population capable of degrading benzyl alcohol increased. As expected, the total number of benzyl alcohol-degrading microorganisms and the optical density of the culture increased as the BOD increased and the benzyl alcohol concentration decreased. When the population approached steady state the number of benzyl alcohol-degrading microorganisms had increased to greater than 10^{12} cfu ml^{-1} (Figure 7), an observation clearly reflected in the BOD. These data demonstrate the utility of BOD for on-line real-time monitoring of the status of a microbial discovery process. Unlike the BOD, both analysis of residual benzyl alcohol concentration by gas chromatography and off-line measurement of biomass

concentration take time and do not provide an instant assessment of the activity of a microbial population.

Using an experimental procedure which enabled the development of a microbial culture growing exclusively on glucose and the introduction of a population change that could be readily characterised, on-line BOD and off-line data from conventional analytical techniques were compared. Comparison of the BOD with off-line analyses such as optical density, viable count and residual substrate concentration clearly demonstrated that the changes observed in the off-line data were also observed in the BOD. The inverse relationship between BOD and residual substrate concentration that was demonstrated in batch culture was also observed in the continuous system. These data demonstrate the utility of BOD for use in real-time monitoring of the effect of changes to a microbial population growing in continuous culture.

EXAMPLE 3 USE OF BOD FOR MICROBIAL DISCOVERY

Discovery of 1-methyl-2-pyrrolidinone-utilising microorganisms

Discovery of 1-methyl-2-pyrrolidinone-utilising microorganisms was performed using the method of the invention by imposing selective pressure (in this case the ability to utilise 1-methyl-2-pyrrolidinone as a sole source of organic carbon and energy) in unison with BOD. The method was performed in the apparatus of Figures 1 and 2. A population of microorganisms with the required characteristics was readily established.

Fresh activated sludge sourced from a wastewater treatment facility was used as the source of microorganisms for enrichment of 1-methyl-2-pyrrolidinone-utilising microbes. As 1-methyl-2-pyrrolidinone is soluble in water it was added to the feed fluid at the 1 g l^{-1} concentration. The enrichment process was performed at 30°C and pH 7.0 (the pH was maintained at 7.0 by the

automatic addition of a potassium hydroxide or hydrochloric acid solution as the alkali and acid, respectively). The feed flow rate was 60 ml h^{-1}.

After the addition of the activated sludge to a vessel the BOD was high (greater than 500 mg l^{-1}). The activated sludge had a high initial BOD because it contained residual readily biodegradable carbon which was gradually degraded, resulting in the observed gradual decline in BOD before the addition of 1-methyl-2-pyrrolidinone. After 2 ml of 1-methyl-2-pyrrolidinone was added to the vessel (Arrow A; Figure 8) a rapid rise in BOD was observed indicating exponential growth (Figure 8). This data can be used to calculate μ_{max} (maximum doubling time) for the population that is growing on the substrate. Growth is exponential between 1220 and 1460 minutes and μ_{max} can therefore be calculated as 0.52 h^{-1}, which corresponds to a doubling time of 1.34 hours. The rapid decline in BOD (Arrow B, Figure 8) was due to the oxygen consumption of the microbial population being greater than the amount of oxygen supplied to the culture.

After initial batch operation, the system was operated in continuous mode (Figure 9) and the feed fluid was increased to a flow rate of 60 ml h^{-1}. It should be noted that when the feed was started the BOD appears to be very low. This is not a true reflection of the status of the culture; the BOD was in fact off-scale (too high) and could not be measured accurately. After the feed pump was started a second exponential rise in BOD was observed which could be attributed to unbalanced growth. The culture will take a period of time to adjust to the fluid flow rate with the usual result being a build-up of the limiting nutrient that is then rapidly depleted as all nutrients are once again in excess. The rapid decrease in BOD after the exponential rise signifies depletion of excess 1-methyl-2-pyrrolidinone as the system approaches equilibrium. The BOD then stabilised at approximately

800 mg l^{-1}, which is the expected value from a feed containing 1.0 g l^{-1} 1-methyl-2-pyrrolidinone (see calculation below):

Balance the stoichiometry of the following equation:

$$C_5H_9ON + O_2 \rightarrow CO_2 + H_2O + NH_3$$

i.e.

$$4C_5H_9ON + 27O_2 \rightarrow 20CO_2 + 12H_2O + 4NH_3$$

Therefore:

Complete oxidation of 4 mol of $C_5H_9NO_2$ requires 27 mol of O_2

Convert from moles to grams:

99.13 x 4 grams of $C_5H_9NO_2$ requires 32 x 27 grams O_2

396.5 g of $C_5H_9NO_2$ requires 864 g of O_2

Concentration of 1-methyl-2-pyrrolidinone in the feed = 1 g l^{-1}, therefore:

1 g of $C_5H_9NO_2$ requires 2.18 g of O_2

Therefore the Chemical Oxygen Demand (COD):

COD = 2180 mg l^{-1}

The BOD is assumed to be one third of the COD:

BOD = 726 mg l^{-1}

The calculated BOD for 1 g l^{-1} 1-methyl-2-pyrrolidinone is less than the measured BOD output. Once

again, the difference is probably due to background respiration. As expected, when the flow of the feed fluid was reduced to 0 ml h^{-1} the BOD dropped rapidly and remained constant at approximately 80 to 120 mg l^{-1}. This background respiration needs to be subtracted from the measured BOD output to give a true indication of the BOD and therefore the measured and calculated BOD are approximately the same. The absolute BOD is not critical for the success of the method of the invention. For microbial discovery the relative value gives a better reflection of the status of a discovery process. For example, the large peak in BOD at the start of the experiment (Figure 9) gives a clear indication of microbial attack of the substrate. The calculated BOD can be used as a guide to select substrate concentrations and other operating parameters. For example by calculating the BOD of a particular substrate the operator can ensure that the substrate concentration in the feed does not exceed the measurable BOD output.

After 116 hours the feed flow rate was increased to 120 ml h^{-1} and shortly after the 1-methyl-2-pyrrolidinone concentration in the feed was increased to 2 g l^{-1} (data not shown). This was continued for a further 95 hours after which a sample was taken for isolation of pure cultures of the microorganisms that were present in the culture. The sample was heavily aggregated with large flocs present and microscopic examination revealed a culture that was dominated by a non-motile rod with a low number of motile rods also being present. The sample was plated onto solid defined medium with 1-methyl-2-pyrrolidinone as the sole carbon source and the plates incubated at 30°C for ~40 hours. From these plates three isolates, designated 2A, 2B and 2C, were purified. Based on microscopic appearance and colonial morphology 2A and 2C were assumed to be the same organism and 2C was pursued no further.

The characteristics of the pure isolates are shown in table 2:

Table 2: Colony morphology and microscopic characteristics of the 1-methyl-2-pyrrolidinone-degrading isolates designated 2A and 2B.

Isolate 2A		Isolate 2B	
Colonial morphology	Microscopic appearance	Colonial morphology	Microscopic appearance
Mucoid	Rod shape	Slightly mucoid	Slightly bent, possibly cocci in chains
Semi opaque	Slight motility	Yellow/white	Non motile
Off white/grey colour	Short rods	1 mm diameter	Cocco-bacilli
2-4 mm diameter	Gram negative	Round colonies	Gram positive
Slimy		Apparent fluorescence	
Apparent fluorescence			

The ability of the pure isolates to grow on 1-methyl-2-pyrrolidinone as the sole source of carbon in liquid culture was also evaluated (Table 3). The cultures were grown in 50 ml screw-capped plastic tubes that contained 10 ml of defined medium and 1.0 g l^{-1} 1-methyl-2-pyrrolidinone. To ensure each culture was inoculated with a consistent number of cells, 10 ml of medium was seeded with 100 µl of a single colony that had been resuspended in 1 ml of DM. The cultures were incubated at 30°C shaking at 190 rpm. A single 10 ml culture was harvested by centrifugation at each time point and the

supernatant kept for determination of the 1-methyl-2-pyrrolidinone concentration. The 1-methyl-2-pyrrolidinone concentrations were estimated using gas chromatography

Table 3: 1-methyl-2-pyrrolidinone degradation by isolates 2A and 2B.

Time after inoculation (h)	Residual 1-methyl-2-pyrrolidinone concentration (mg l^{-1}) Uninoculated Control	Isolate 2A	Isolate 2B
24	1000	ND[a]	ND
48	960	ND	ND
72	850	ND	ND
96	870	ND	ND
168	900	ND	ND

a Not detected (limit of detection = 20 mg l^{-1}).

The results show that from a large mixed population (activated sludge) two isolates were obtained that were able to use 1-methyl-2-pyrrolidinone as the sole source of carbon. Both these isolates were able to completely degrade 1.0 g l^{-1} 1-methyl-2-pyrrolidinone in batch culture within 24 hours.

The BOD output demonstrates the usefulness of BOD as a real-time monitor of the status of a culture. Any changes to the operating conditions are reflected almost immediately in the visual output. This enables the operator to make changes and note the response of the culture rapidly without the requirement for off-line analyses which are time consuming and result in a delay before the effect of a change can be assessed. Additionally, growth on 1-methyl-2-pyrrolidinone was

demonstrated without the need for development of an assay for the substrate. This has the added benefit in that insoluble substrates (see Example 4) can be assessed which can be difficult to assay because a representative sample cannot be taken and analysed easily.

EXAMPLE 4 DISCOVERY OF DODECANE-UTILISING MICROORGANISMS

Discovery of dodecane-utilising microorganisms was performed using the method of the invention. By imposing selective pressure (in this case the ability to utilise dodecane as a sole source of carbon and energy) in unison with monitoring the BOD output, a microbial population with the required characteristics was readily established. As dodecane is practically insoluble in water it was fed into the culture using a separate peristaltic pump at a flow rate of 0.79 ml h^{-1}. The purpose of this example was to discover microbes that could hydroxylate linear hydrocarbons. This is extremely difficult to achieve using convention chemical (non-microbial) techniques.

Fresh activated sludge sourced from a wastewater treatment facility was used as source of microorganisms for discovery of dodecane-utilising microbes. The process was conducted on the apparatus of Figures 1 and 2. The discovery process was performed at 30°C and pH 7.0 (the pH was maintained at 7.0 by the automatic addition of a potassium hydroxide or hydrochloric acid solution). The feed was comprised of DM that had no carbon source added and the feed flow rate was initially 30 ml h^{-1} and the flow of dodecane was 0.79 ml h^{-1}. The experiment was conducted over 137 hours then the apparatus components cleaned and restarted (using the same culture) (*Arrow B*) with a feed flow rate of 60 ml h^{-1} (the dodecane flow rate was unchanged). After 330 hours of operation the fluid was sampled to enable isolation of dodecane-degrading microorganisms (*Arrow C*, Figure 10).

Although the BOD output was variable, clearly a population of dodecane-degrading microorganisms had been established. The population took somewhat longer to establish than was observed with the water soluble substrate 1-methyl-2-pyrrolidinone. Two possible explanations for this observation are (i) the variability of substrate flow resulted in gradual washout of any dodecane-degrading population that may have become established and/or (ii) the insoluble nature of the substrate reduces microbial attack resulting in slower growth. A combination of gradual washout and an insoluble substrate could result in reduced degradation of the substrate because the enriched population may be producing surfactants or similar molecules that assist in solubilising the substrate. Gradual washout would continually reduce the concentration of any surfactant-type molecules further decreasing the accessibility of the substrate resulting in a continual compounding negative effect. In this experiment dodecane was fed into the reactor using a peristaltic pump which resulted in the variable BOD output. Syringe pumps or peristaltic pumps can be used to feed insoluble substrates into the culture, however a syringe pump is preferred because the product contact components of a syringe pump are compatible with a wide range of chemicals.

The BOD output is significantly less than the calculated value based on the COD of dodecane (see calculation below).

Balance the stoichiometry of the following equation:

$$C_{12}H_{26} + O_2 \rightarrow CO_2 + H_2O$$

i.e.

$$2C_{12}H_{26} + 37O_2 \rightarrow 24CO_2 + 26H_2O$$

Therefore, complete oxidation of 2 mol of $C_{12}H_{26}$ requires 37 mol of O_2

Convert from moles to grams:

170.3 x 2 grams of $C_{12}H_{26}$ requires 32 x 37 grams O_2

340.6 g of $C_{12}H_{26}$ requires 1184 g of O_2

Assuming the concentration of dodecane in the feed = 1 g l^{-1}, therefore:

1 g of $C_{12}H_{26}$ requires 3.48 g of O_2

Therefore the Chemical Oxygen Demand (COD):

COD = 3480 mg l^{-1}

The BOD is assumed to be one third of the COD:

BOD = 1158 mg l^{-1}

Actual flow rate of dodecane = 0.788 ml h^{-1} = 0.591 g h^{-1}

Therefore the estimated dodecane concentration in feed = 0.591/60ml = 9.85 g l^{-1}

Expected COD = 28861 mg l^{-1} and BOD = 9620 mg l^{-1}

This result was unexpected although it could be explained by the insolubility of the substrate. As dodecane is less dense than water it will tend to float to the surface of the culture, particularly during the measurement of oxygen uptake where aeration has stopped and the stirrer has slowed. Much of the dodecane may be washed out in the overflow. The measured BOD may also be an indication of the amount of substrate that is accessible to the microbial population and that is limited by the solubility of dodecane in water.

After 207 hours of growth on dodecane a sample was taken from the enriched culture for isolation of pure cultures. Microscopic examination of the sample revealed a range of rod-shaped bacteria both short and filamentous. Cocci-shaped bacteria were also evident and a number of motile rods were also observed. The sample was plated onto solid DM with dodecane as the sole carbon source and the plates were incubated at 30°C for ~48 hours. From these plates four isolates, designated 1A, 1B, 1C and 1D, were purified. Based on microscopic appearance and colonial morphology 1A and 1D were assumed to be the same organism and 1A was pursued no further.

The characteristics of the pure isolates are shown in table 4:

Table 4: Colony morphology and microscopic characteristics of the dodecane-degrading isolates designated 1B, 1C and 1D.

Isolate 1B		Isolate 1C		Isolate 1D	
Colonial morphology	Microscopic appearance	Colonial morphology	Microscopic appearance	Colonial morphology	Microscopic appearance
Shiny	Cocco-bacilli	Fried egg appearance	Long and short rods	Small uneven colonies	Long rods
Round	Non-motile	Target shaped	Motile	Crinkly Appearance	Non-motile
Off white colour	Gram negative	3-6 mm diameter	Gram negative	Off white colour	Gram negative
~1.5 mm diameter		Apparent fluorescent halo		Hazy/opaque appearance	

The ability of the pure isolates to grow on dodecane as the sole source of carbon in liquid culture was also evaluated and is shown in Table 5. The cultures were grown in 50 ml screw-capped plastic tubes that contained 10 ml of defined medium and 0.75 g l^{-1} dodecane. To ensure each culture was inoculated with a consistent number of cells, 10 ml of medium was inoculated with 100 µl of a single colony that had been resuspended in 1 ml of DM. The cultures were incubated at 30°C shaking at 190 rpm. Residual dodecane was extracted by the addition of 20 ml of hexane to a single 10 ml culture at each time point. The tube was shaken vigorously for one minute and after phase separation, the upper layer was kept for determination of the dodecane concentration. The dodecane concentrations were estimated using gas chromatography.

Table 5: Dodecane degradation by isolates 1B, 1C and 1D.

Time after inoculation (h)	Residual dodecane concentration (mg l^{-1}) Uninoculated Control	Isolate 1B	Isolate 1C	Isolate 1D
24	820	920	630	1080
48	800	160	620	920
72	200	420	440	740
96	740	270	440	410
168	740	290	280	310

The results show that from a large mixed population (activated sludge) three isolates were obtained that were able to use dodecane as the sole source of carbon. In batch culture the isolates were able to use (over a 168 hours period) 50 to 60% of the dodecane added to the culture. The rate of dodecane utilisation is substantially slower than 1-methyl-2-pyrrolidinone which may be due to the difference in the solubility of the two compounds (dodecane is practically insoluble in water). The insolubility of dodecane may impose mass transfer limitations which will slow growth and utilisation of the substrate significantly. The variability in the gas chromatography data from the batch experiments highlights the difficulties associated with analysis of concentrations of insoluble substrates. This problem can be partly overcome by monitoring BOD, as oxygen consumption can be used as an indirect indicator of growth on the substrate.

EXAMPLE 5 DISCOVERY OF OLIVE OIL-UTILISING MICROORGANISMS

The use of olive oil as a feed fluid highlights another of the advantages of the method of the invention, which is microbial discovery in extreme environments. Olive oil is a heterogeneous substrate of which development of an analytical method for measuring consumption would be difficult. Monitoring BOD enables demonstration of growth on this complex substrate without the requirement for the development of complex analytical methods. The isolation of microorganisms capable of using substrates such as olive oil for growth may enable the discovery of lipases with useful properties. The following experiment was performed to facilitate not only the isolation of olive oil-degrading microorganisms but also to enrich microbes that can tolerate a very broad pH range.

The vessel was filled with activated sludge and 10 ml of olive oil was added. The BOD rose rapidly and peaked at ~1700 mg l^{-1}. The rapid onset in the ability to degrade olive oil in a population of microorganisms from activated sludge is not unexpected as the presence of this type of substrate in the influent streams of wastewater treatment facilities is highly likely. After the peak in BOD was observed (20.5 hours) olive oil was fed continuously into the vessel as was a separate stream from DM mixed with activated sludge in the ratio 4:1. The pH set point was reduced to pH 4.0 and after 225 hours the feed medium was changed from a mixture of DM and activated sludge to DM. No changes were made to the conditions for 138 hours (equivalent to 11 vessel volumes) and the BOD of the culture remained high. From these observations it was concluded that a microbial population had been established that could use olive oil as the sole source of carbon at pH 4.0.

The olive oil flow rate was reduced to 0.061 ml h^{-1} and the feed was again changed to a mixture of activated

sludge and DM. These conditions resulted in a culture with activity (BOD in the range 1200 to 1500 mg l^{-1}) then after 458 hours the pH set point was changed to pH 2.2, the feed was again changed to DM without any additions and the feed flow rate was increased to 66 ml h^{-1}. The conditions were unchanged for 55 hours (4.4 vessel volumes) and the BOD stabilised at ~1700 mg l^{-1} indicating that a population of microorganisms had been established that was capable of using olive oil as a sole source of carbon at pH 2.2.

The next phase of the experiment evaluated the ability of the microbial population that had been growing at pH 2.2 to respond to an increase in the pH of the culture. At 555 hours the pH set point was increased to 9.0. A further increase in the pH set point from pH 9.5 to 10 resulted in another decline in BOD indicating washout and/or death of the microbial population. Interestingly, when the pH was reduced by just 0.5 of a pH unit the culture recovered with the BOD increasing exponentially. The culture showed significant sensitivity to pH values greater than 9.5. The reason for this observation is not clear however two possible explanations for the increased sensitivity to pH 10 could be (i) one of the medium components was insoluble at pH 10 resulting in significant nutrient limitation and a decline in BOD or (ii) the microbial population present in the culture had not adapted to growth at pH 10. The culture was maintained at pH 9.5 for 125 hours and clearly a population of microorganisms growing on olive oil as the sole carbon source at pH 9.5 had been established. It cannot be concluded that this population also has the potential to grow at pH 2 because the time taken to establish this population may have resulted in the development of a totally new population that is better adapted to growth under the new conditions.

Over a 37 day period growth of a microbial population on olive oil at a range of pH values was demonstrated. The two extremes of pH were 2.2 and 9.5. Clearly microbial

activity could be demonstrated at these pH values and these data were then used to develop an automatic pH oscillation system; this system was designed to facilitate the isolation of microorganisms with tolerance to a broad pH range with the view to isolating enzymes from these microorganisms which exhibit similar pH tolerance (both activity and stability).

EXAMPLE 6 DEVELOPMENT OF A FEEDBACK LOOP BETWEEN FEED FLOW RATE AND OUR

A feedback loop between feed flow rate and OUR was developed to enable the maximum growth rate of a microbial population to be established using an automated system. The maximum growth rate of a population is an important parameter as this is likely to give an indication of the rate of flux through a metabolic pathway.

6.1 Design of feedback loop

The feedback loop uses the limiter that if the BOD remains within a set range for a (operator set) period then the flow rate is increased by a value that is also specified by the operator. This is described briefly above in relation to the apparatus of the embodiment illustrated in Figures 1 and 2. The software to run the feedback loop was developed using VisiDAQ Builder and Adam A/D modules were used for signal processing. The notable components of the system are represented in Figure 11 as the "Flow Calibration" (Arrow A) and "BOD Control" (Arrow B).

6.2 Testing of the feedback loop between feed flow rate and OUR

Isolation of 1,3-propanediol-degrading microorganisms was used to test the feedback loop between feed flow rate and OUR. The feed medium was a defined medium designated 461S (Appendix I) which contained 1.0 g l^{-1} 1,3-propanediol and the initial flow rate was 43.5 ml h^{-1}. The operating temperature was 30°C and pH 7.0. The medium was inoculated with ~700 ml of activated sludge. The feed flow rate was

increased by 20 ml h^{-1} if the BOD remained constant for four vessel volumes. After an initial peak in the BOD, which was due to 1,3-propanediol being in excess, the BOD remained constant over a range of flow rates. The flow rate was increased in steps from 43.5 ml h^{-1} to 143.5 ml h^{-1} over several days without any significant change in the BOD, demonstrating that the microbial population which had been established was capable of growing with doubling times in the range 3.6 to 12 hours. At a doubling time of 3.6 hours the BOD was unchanged suggesting that the population that had been established had the ability to grow faster than the maximum that was tested in this experiment. This observation was expected because the inferred μ_{max} of the population from the initial peak in BOD at the start of the experiment was ~0.25 h^{-1} (a doubling time of 2.8 hours). Higher doubling times could be achieved with any microbial population that is established in the culture because there is a good probability that mutants which can grow at a higher rate will be selected at high feed flow rates.

The response of the culture to the changing flow rates is shown in Figure 12.

Modifications may be made to the preferred embodiments and Examples described above without departing from the spirit and scope of the invention.

Appendix I

Media

Composition of Defined Medium (DM)

	g l^{-1}
NH_4Cl	1.0
KH_2PO_4	0.5
10% Na_2SO_4	2.0 ml l^{-1}
*$MgCl_2.6H_2O$	0.17
*$CaCl_2.2H_2O$	0.01
**Trace Metals solution	1.0 ml l^{-1}

All media were made up in reverse osmosis water and adjusted to pH 7.0 with 4M NaOH

All chemicals were of analytical grade.

Where required, media were sterilised by autoclaving at 121°C for 20 minutes. Large volumes (up to 20 litres) of feed were autoclaved at 121°C for at least 60 minutes.

* Magnesium and calcium were added as a concentrated sterile stock solution (17.0 g l^{-1} $MgCl_2.6H_2O$; 1.0 g l^{-1} $CaCl_2.2H_2O$) after autoclaving to prevent precipitation with orthophosphate.

Carbon sources were added after the media were autoclaved.

Solid media were prepared by the addition of 15 g l^{-1} agar.

** The Trace Metals solution contained:

	g l^{-1}
$FeSO_4.7H_2O$	1.0
$CoSO_4.7H_2O$	0.2
$MnSO_4.H_2O$	0.1
$NiCl_2.6H_2O$	0.1
$NaMoO_4.2H_2O$	0.05
H_3BO_3	0.062
$ZnCl_2$	0.07
$CuSO_4.5H_2O$	0.02

Composition of Defined Medium (461S) which is a modification of a minimal medium described by Nagel and Andreesen as cited by DSMZ (German culture collection – www.dsmz.de/media).

	ml l^{-1}
*Salts solution	10
**Trace Elements Stock	0.7
***Phosphates	20

* The Salts solution contained:

	g l^{-1}
$CaCl_2.2H_2O$	1.0
$MgSO_4.7H_2O$	50.0
$MnSO_4$	1.0
NH_4Cl	30.0
NaCl	5.0

** The chemicals in the Trace Elements Stock were dissolved 5M HCl. The Trace Elements Stock contained:

(Note: $FeSO_4.7H_2O$ was dissolved in the 5M HCl before the addition of the other components.)

	g l^{-1} (of 5M HCl)
$FeSO_4.7H_2O$	6.56
$ZnCl_2$	0.14
$MnSO_4.H_2O$	0.12
H_3BO_3	0.01
$CoSO_4.7H_2O$	0.45
$CuSO_4.5H_2O$	0.004
$NiCl_2.6H_2O$	0.048
$NaMoO_4.2H_2O$	0.072

*** The Phosphates solution contained:

	g l^{-1}
Na_2HPO_4	72.5
KH_2PO_4	12.5

All media were made up in reverse osmosis water and all chemicals were of analytical grade.

Media were prepared by the mixing the Salts solution and the Trace Elements Stock prior to autoclaving.

Where required, media were sterilised by autoclaving at 121°C for 20 minutes. Large volumes (up to 20 litres) of feed were autoclaved at 121°C for at least 60 minutes.

The Phosphates solution was added after autoclaving to prevent precipitation of orthophosphates with the metals in the medium.

Carbon sources were added after the media were autoclaved.

Solid media were prepared by the addition of 15 g l^{-1} agar.

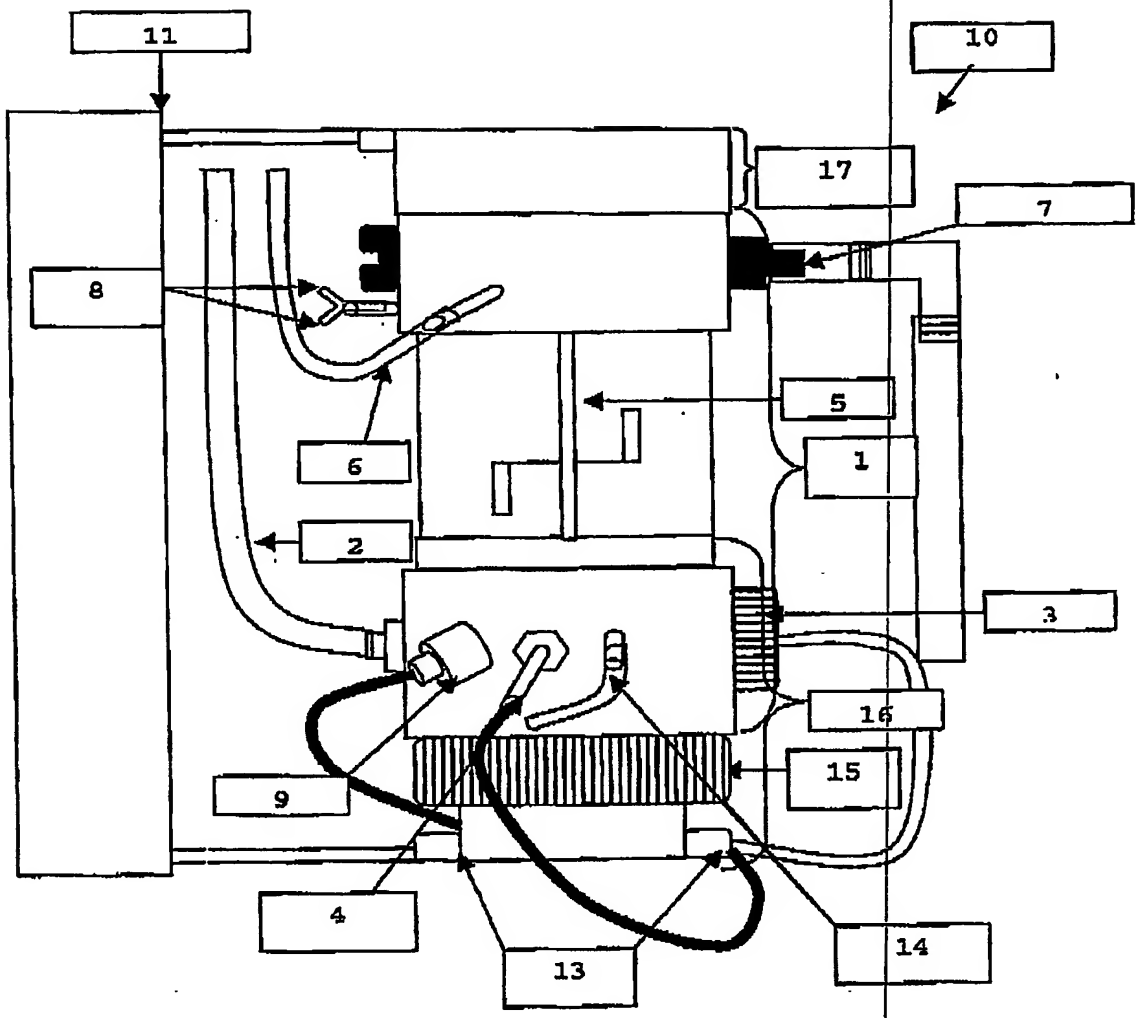

Figure 1

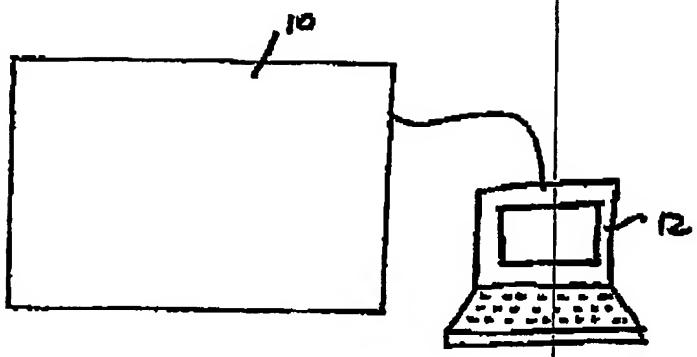

Figure 2

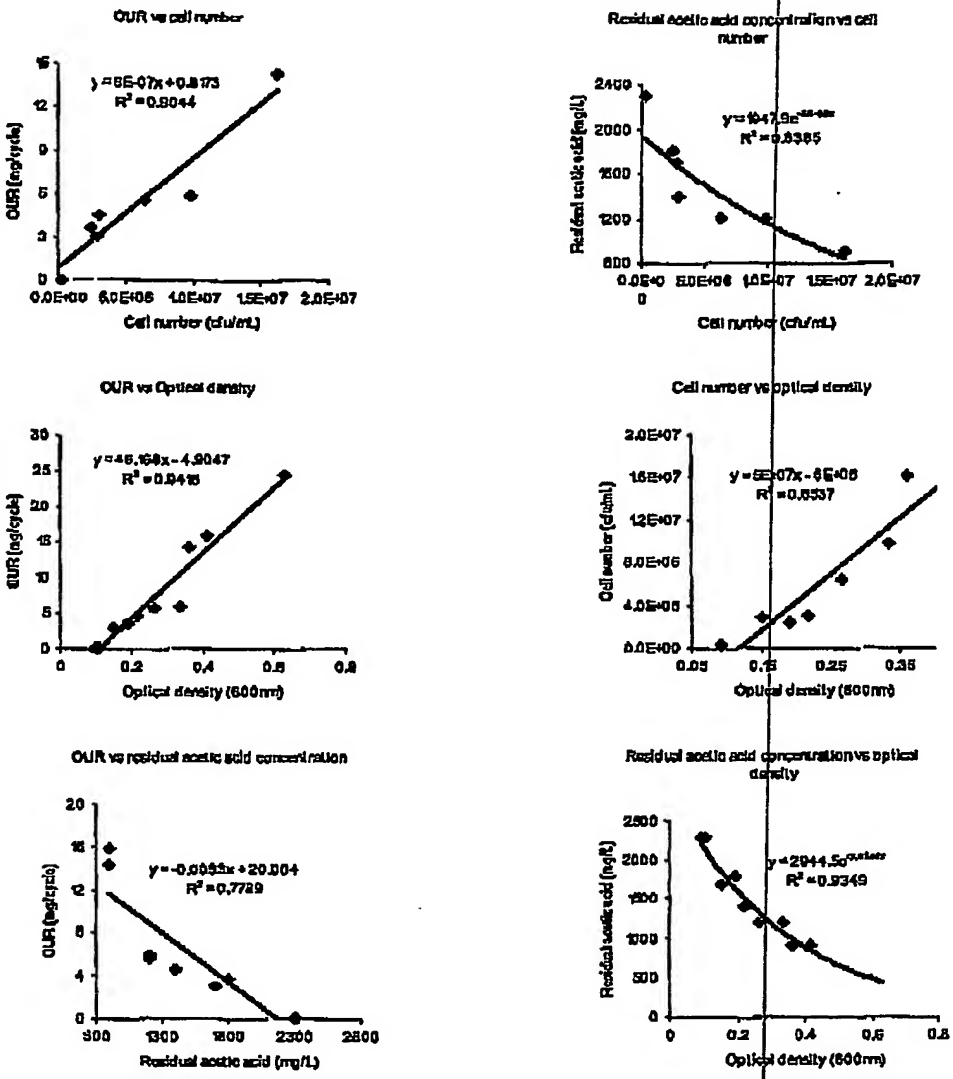

Figure 3

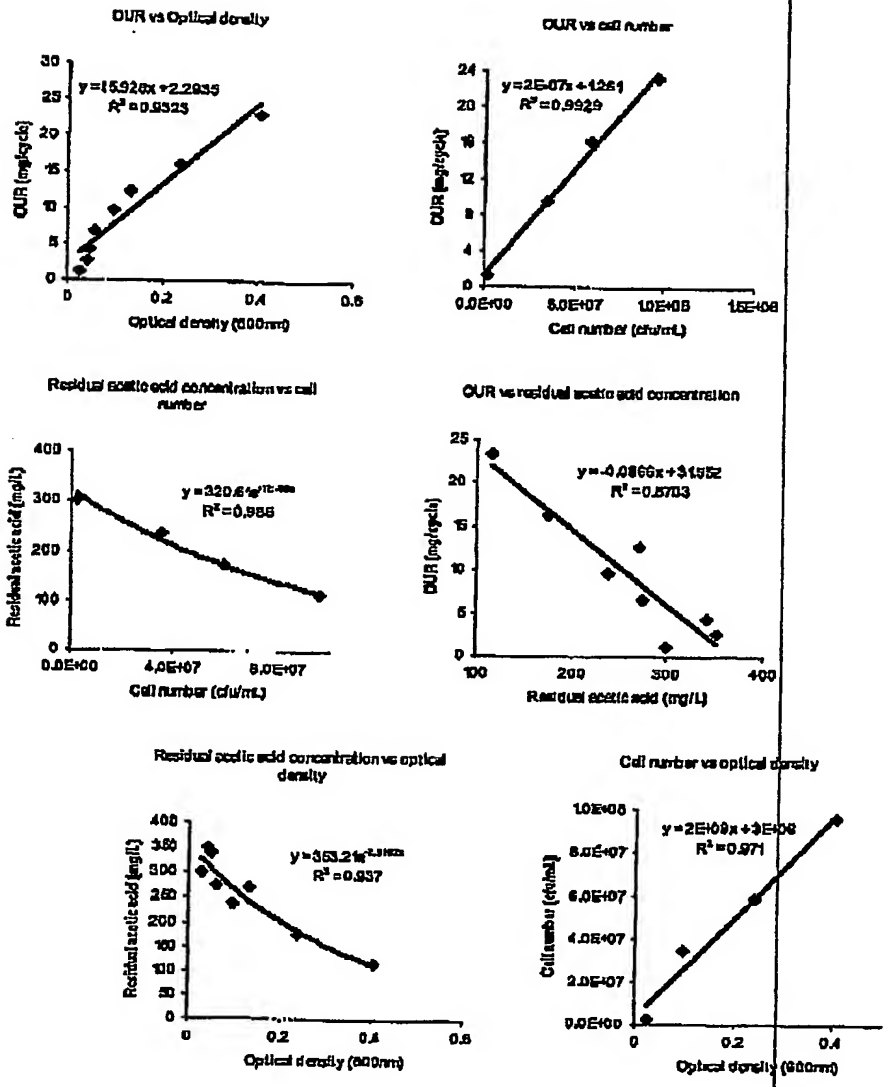

Figure 4

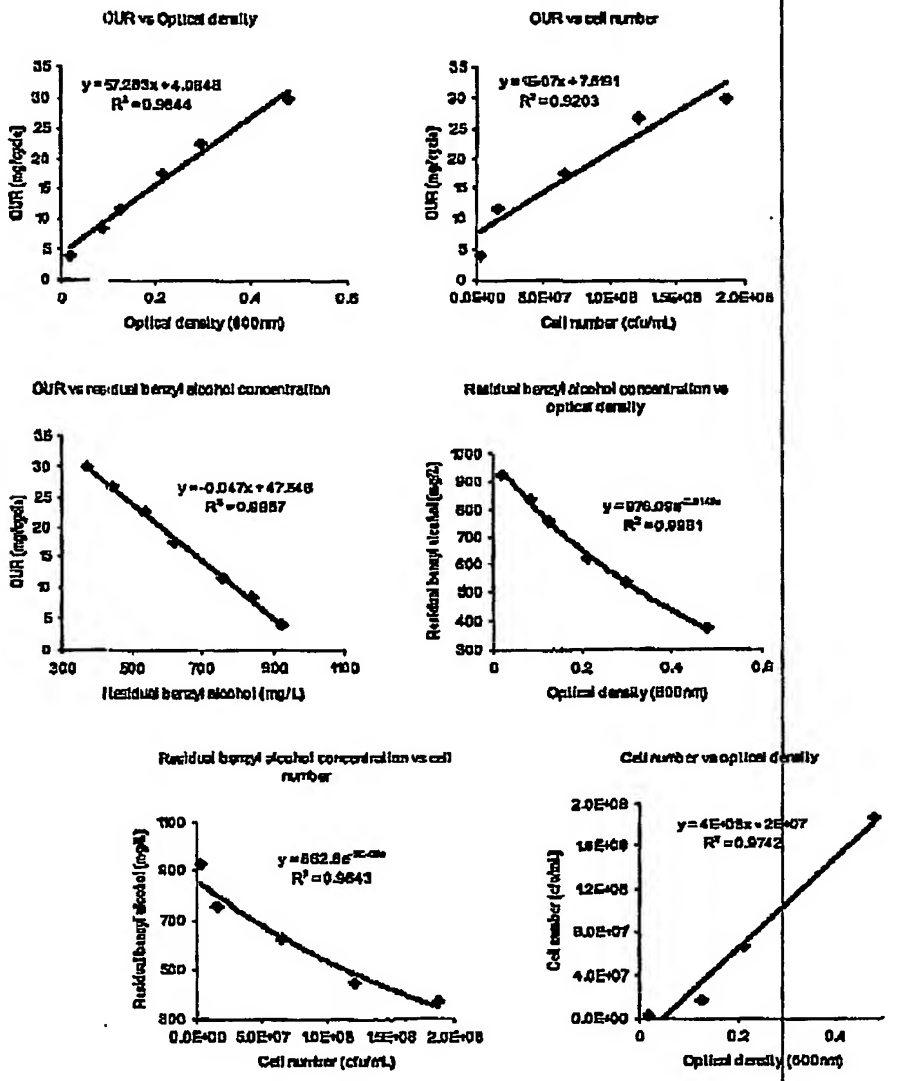

Figure 5

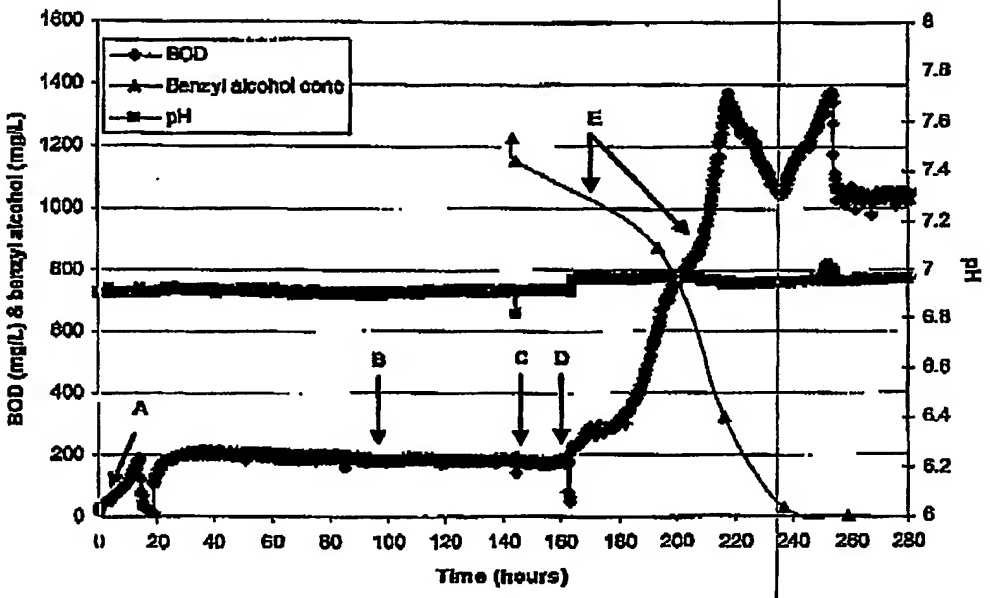

Figure 6

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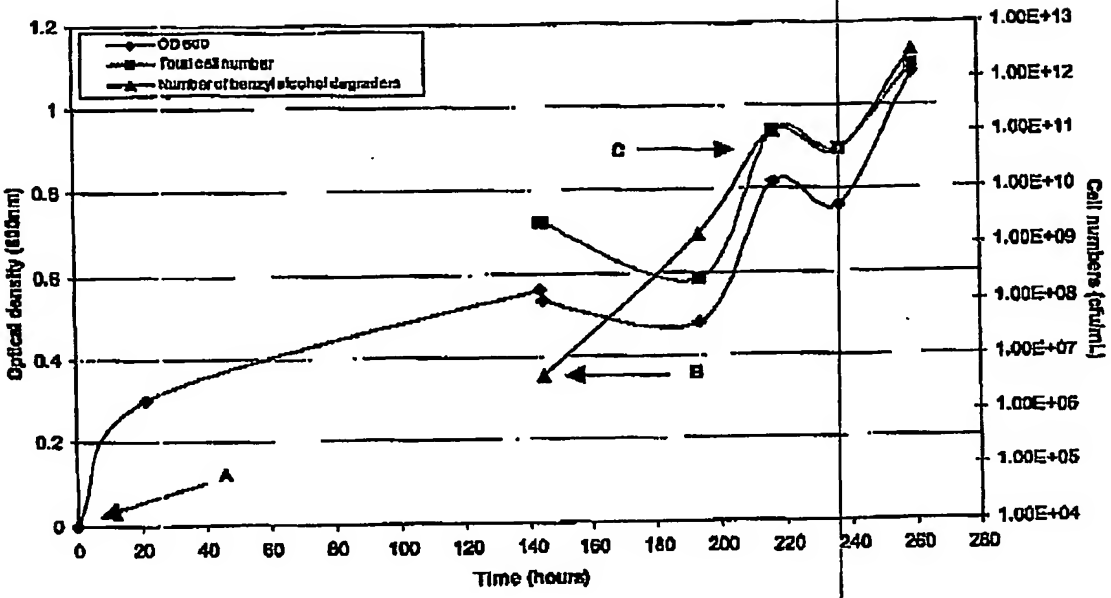

Figure 7

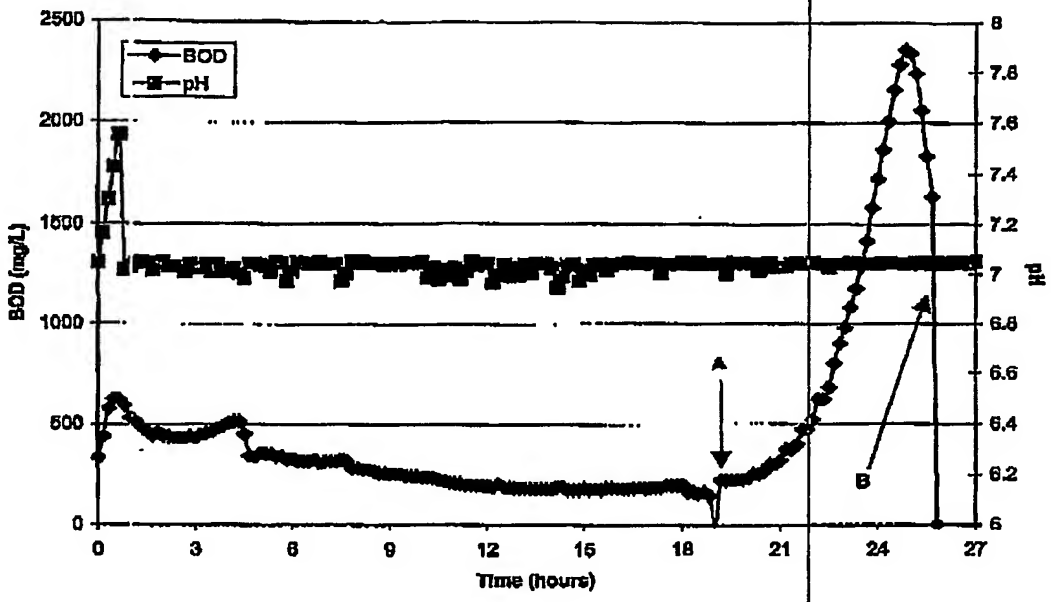

Figure 8

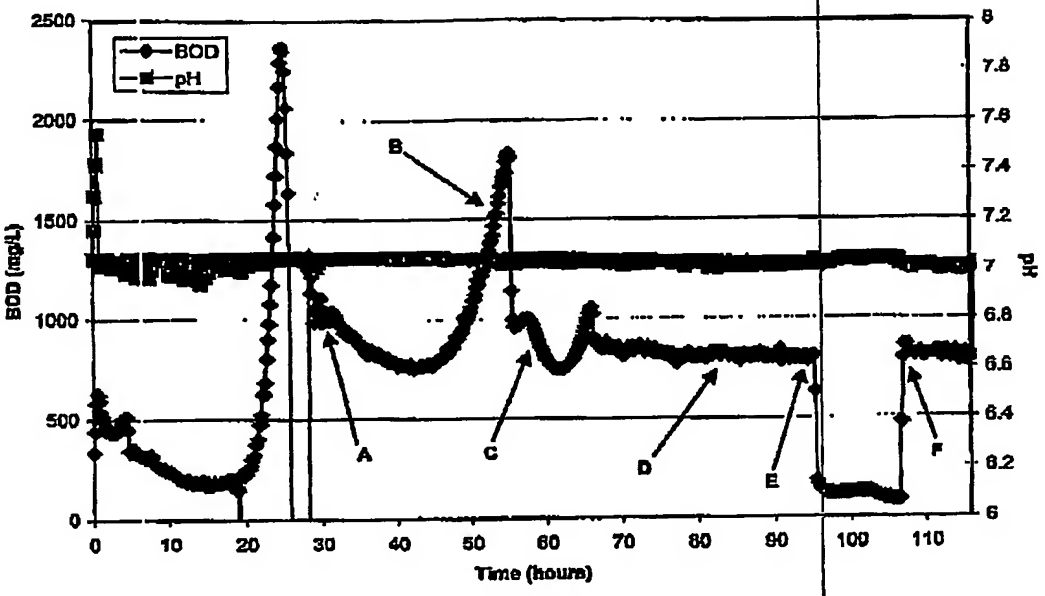

Figure 9

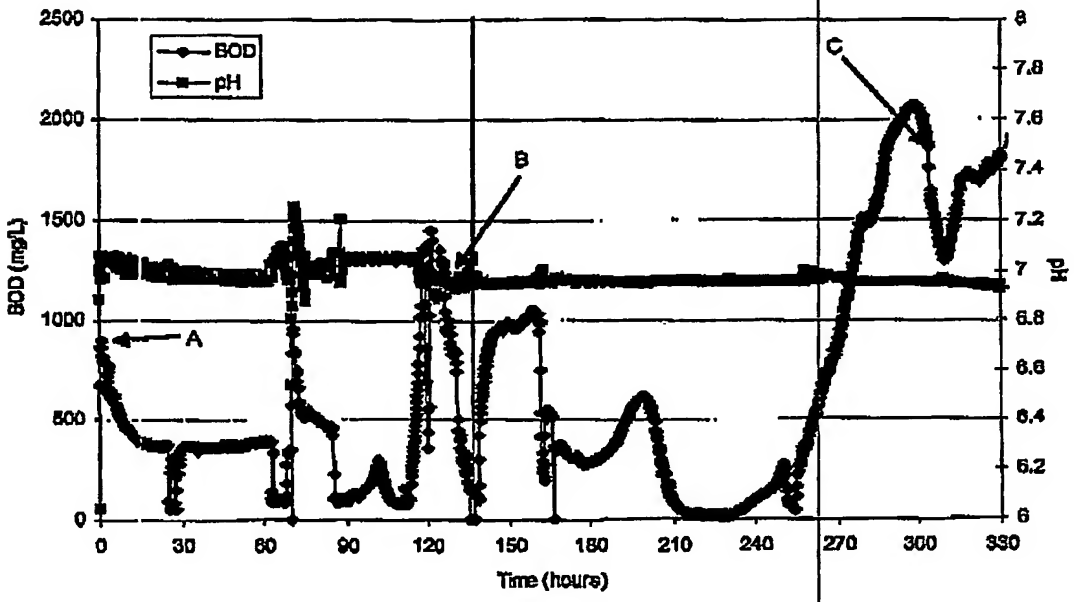

Figure 10

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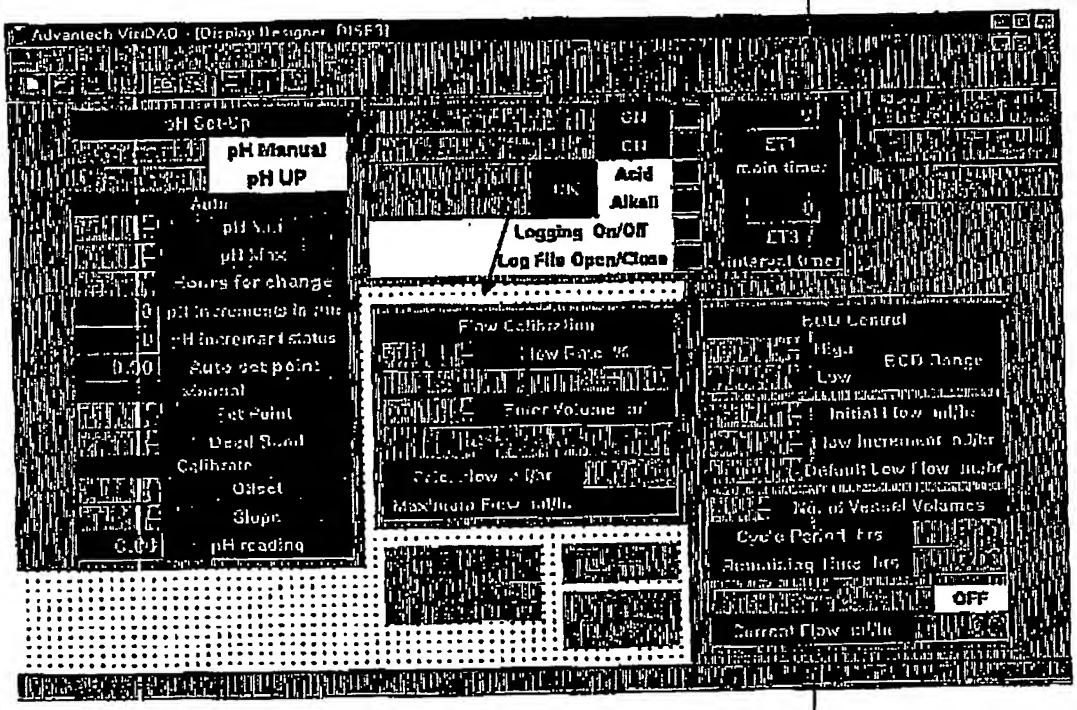

Figure 11

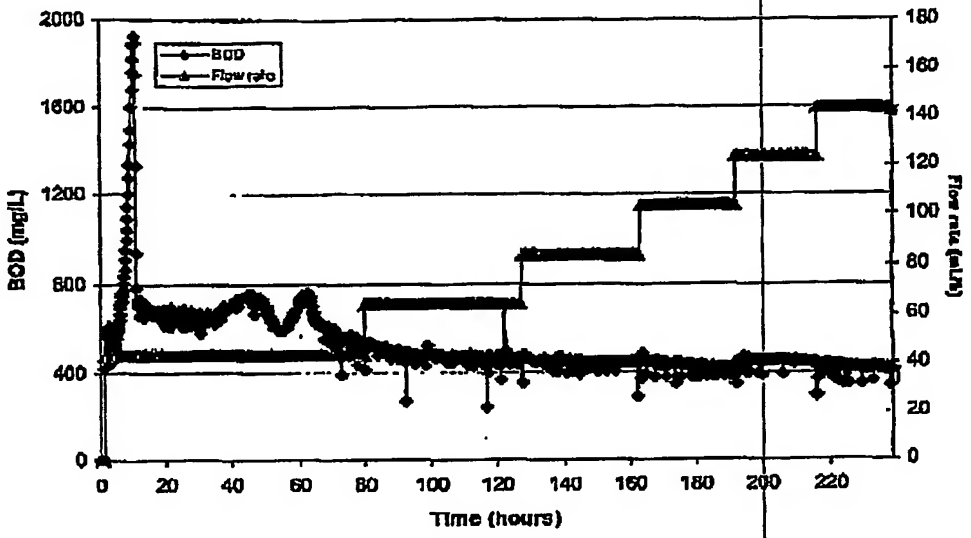

Figure 12

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